

Identification of specific probiotics capable of enhancing resistance to bacterial coldwater disease (CWD) in rainbow trout

Kenneth D. Cain¹, David R. Burbank¹, Scott E. LaPatra²,
and Gary Fornshell³

Northwest Fish Culture Conference, December 6-8, 2011

¹ Department of Fish and Wildlife and Aquaculture Research Institute, University of Idaho, Moscow, ID 83844

²Clear Springs Foods, Inc. Research and Development, Buhl, ID 83316

³University of Idaho, Twin Falls County Extension, Twin Falls, ID 83301

What are probiotics?

- “Microbial cells...with the aim of improving health” (Gatesoupe, 1999)
- Production of inhibitory compounds, competitive exclusion, improving host immunity, enhancing water quality, etc.
- Probiotics for humans and terrestrial animals are well established
 - Relatively new to aquaculture
 - Growing research – “green” approach to disease management



www.keetonaqua.com

Probiotics Background

Current selection methods:

- Probiotic candidates which inhibit specific pathogens
- Characteristics of probiotics
 - Enter/survive the gastrointestinal tract
 - Safe to animal and common in environment
 - Inhibit specific pathogenic bacteria

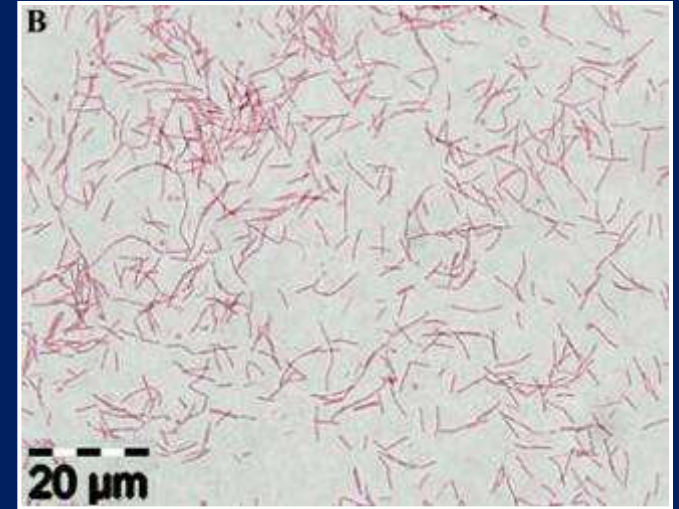
Coldwater Disease

- *Flavobacterium psychrophilum* is causative agent (Rainbow Trout Fry Syndrome in Europe)
- Significant economic losses affecting public and private salmonid aquaculture in the Pacific Northwest and worldwide



Management Options

- Licensed vaccine not yet available
 - Fish must be immunocompetent
- Antibiotics continue to be the main form of treatment
 - Risk of antibiotic resistance?
- Dramatic need for additional options
 - Probiotics as alternative?



Research Objectives

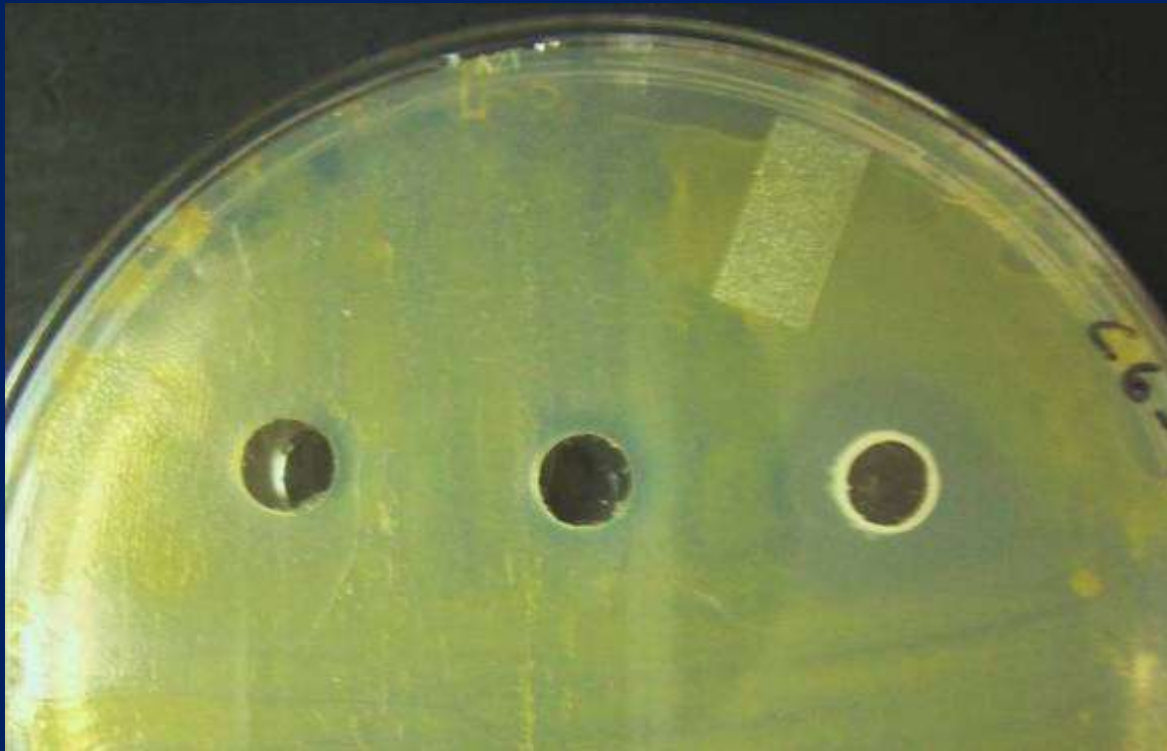
1. Identify candidate probiotics that fit standard definition with ability to inhibit *F. psychrophilum* *in vitro*
2. Test candidate probiotics through feed experiments and measure effects following challenge of fish with *F. psychrophilum*

Objective 1: Isolation of candidate probiotics

- Collected rainbow trout from Idaho
 - Commercial production hatcheries
 - Aquaculture Research Institute (UI)
 - Spring Valley Reservoir (Troy, Idaho)
- Removed gastrointestinal tract, expelled contents, homogenized and plated (TSA, TYES agar)
 - Subcultured until isolation was achieved
- Collected 318 bacterial isolates from 29 rainbow trout

Objective 1: *F. psychrophilum* inhibition *in vitro*

- Out of 318 isolates, 24 were identified which inhibited *F. psychrophilum* growth *in vitro*



Objective 1: Discussion

- Further selection = 16 candidate probiotics
 - Capable of inhibiting *F. psychrophilum* *in vitro*
 - Evidence of survival in the GI tract of rainbow trout *in vitro*
 - Non-pathogenic to rainbow trout

Objectives

1. Identify candidate probiotics that fit standard definition with ability to inhibit *F. psychrophilum* *in vitro*
2. Test candidate probiotics through feed experiments and measure effects following challenge of fish with *F. psychrophilum*

Objective 2: Feed preparation

- Direct feeding of probiotics is the most practical and common method of introduction
- Probiotic feed preparation
 - Candidate probiotic grown to log phase
 - Centrifuge and re-suspend in menhaden oil to $\sim 10^8$ CFU ml⁻¹
 - Top-coated onto commercial trout feed



Objective 2: *In Vivo* Evaluation

- Evaluated the 10 most promising candidate probiotics through four separate trials

	Candidate probiotics
1	<i>Aeromonas caviae</i> ³
2	<i>Aeromonas caviae</i> ⁴
3	<i>Aeromonas caviae</i> ⁵
4	<i>Aeromonas sobria</i> ³
5	<i>Aeromonas sobria</i> ⁴
6	<i>Aeromonas sobria</i> ⁵
7	<i>Aeromonas sobria</i> ⁶
8	<i>Aeromonas sobria</i> ⁷

	Candidate probiotics
9	<i>Lysinibacillus fusiformis</i>
10	<i>Citrobacter freundii</i>
11	<i>Enterobacter</i> sp. ¹ (C6-6)
12	<i>Enterobacter</i> sp. ² (C6-8)
13	<i>Hafnia alvei</i>
14	<i>Plesiomonas shigelloides</i> ¹
15	<i>Plesiomonas shigelloides</i> ²
16	<i>Staphylococcus equorum</i>

Discussion

- Two initial trials conducted: (tested 3 probiotics)
 - *Enterobacter* sp.¹ (C6-6) – appeared promising
 - Adjusted feeding regime for trials 3 and 4
- To observe maximum effect should feed probiotic prior to and during infection
 - Allow more time for colonization and establishment in GI tract



Objective 2: Trial 3 and 4

- Evaluated 7 remaining probiotic treatments in two challenges - also included *Enterobacter* sp.¹ (C6-6)
- Triplicate groups of 25 fish per probiotic treatment
- Fed candidate probiotic for 10 days prior to injection challenge with *F. psychrophilum*
- Continued to feed probiotics throughout 28 day challenge period

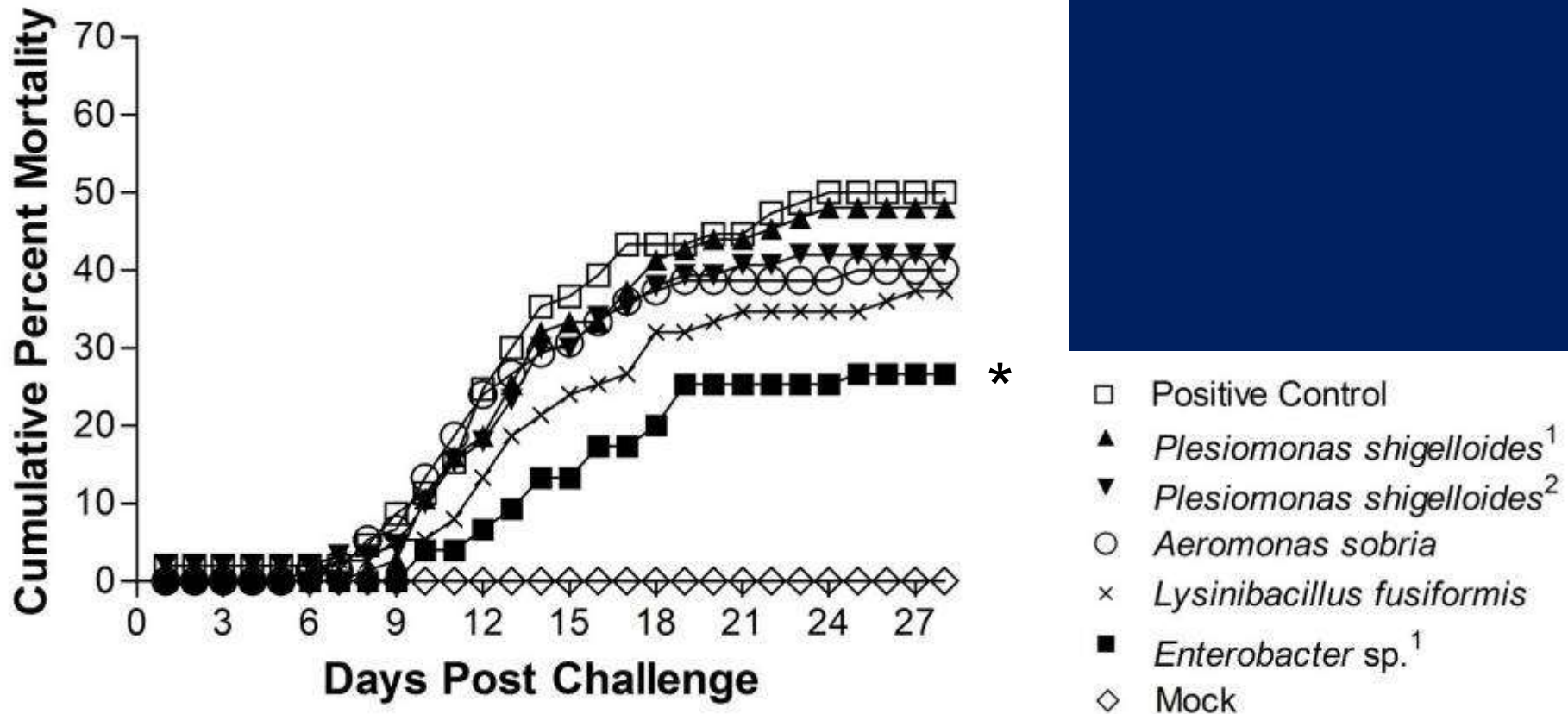


Objective 2: Trial 3 and 4

- Positive control comprised of triplicate groups of 25 fish
 - *F. psychrophilum* injection and feed with oil only
- Mock infected group comprised of triplicate groups of 25 fish - PBS injection and fed standard trout diet
- Monitor mortalities for 28 days
- Attempted to reisolate *F. psychrophilum* and candidate probiotics from at least 20% of daily mortalities

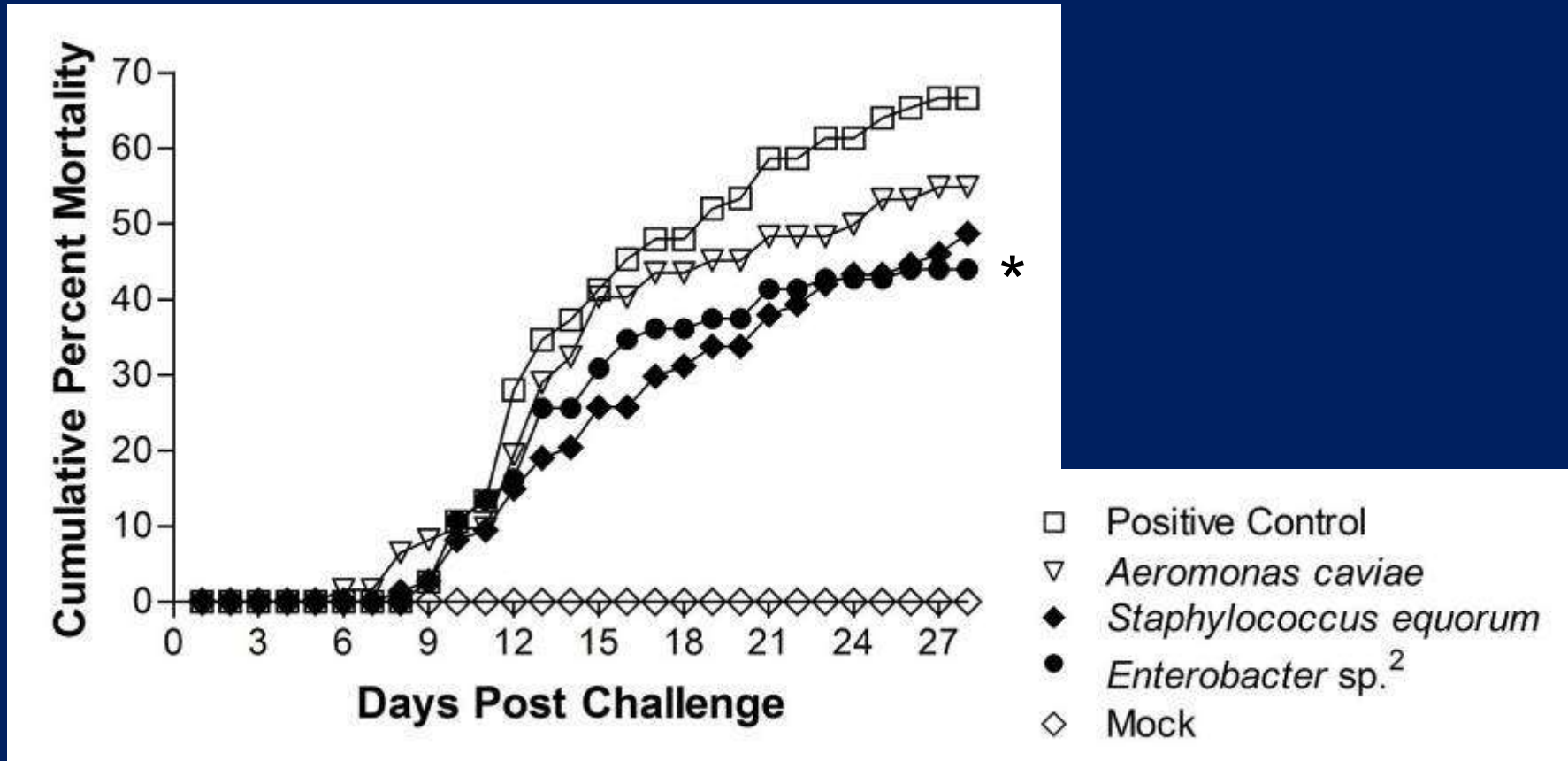


Objective 2: Trial 3



* significant difference (p<0.05) from positive control

Objective 2: Trial 4



* significant difference ($p < 0.05$) from positive control

Objective 2: Discussion

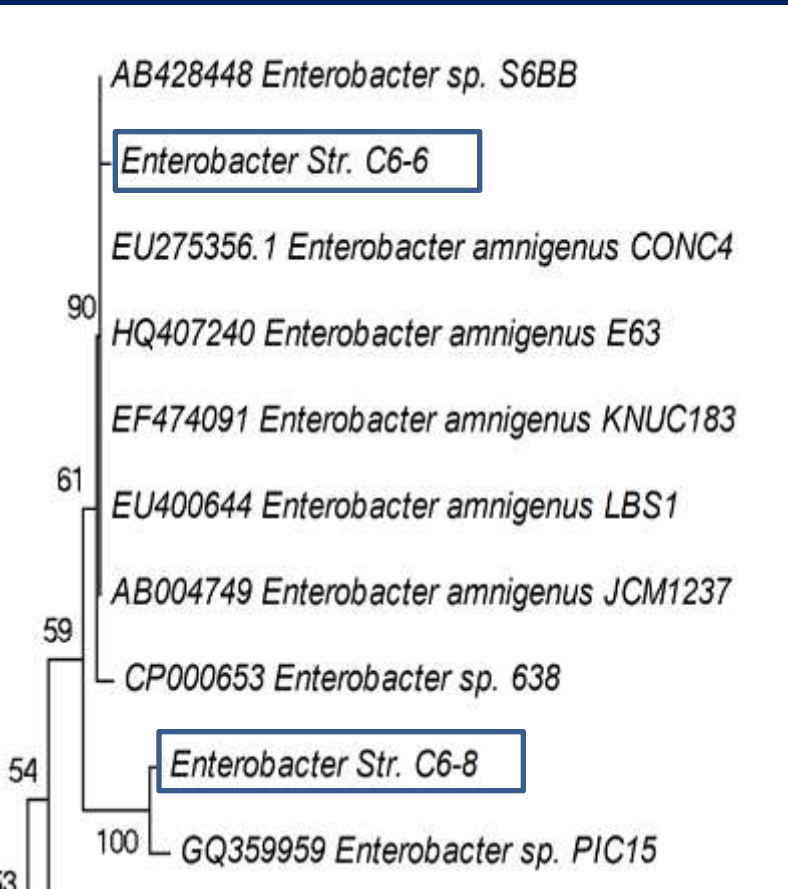
- Both *Enterobacter* sp. (C6-6 and C6-8) are promising probiotics
 - Significant reductions in mortality
 - Consistent reisolation from GI tract compared to other candidates
- All other treatments eliminated from further testing
 - Did not show “significant” decrease in mortality

Enterobacter sp.

- C6-6 and C6-8 believed to be different species based on lab culture characteristics
- 16S sequencing confirmed that these are separate *Enterobacter* species
 - Most closely related with *Enterobacter amnigenus* and *Enterobacter* sp. PIC15

Enterobacter sp.

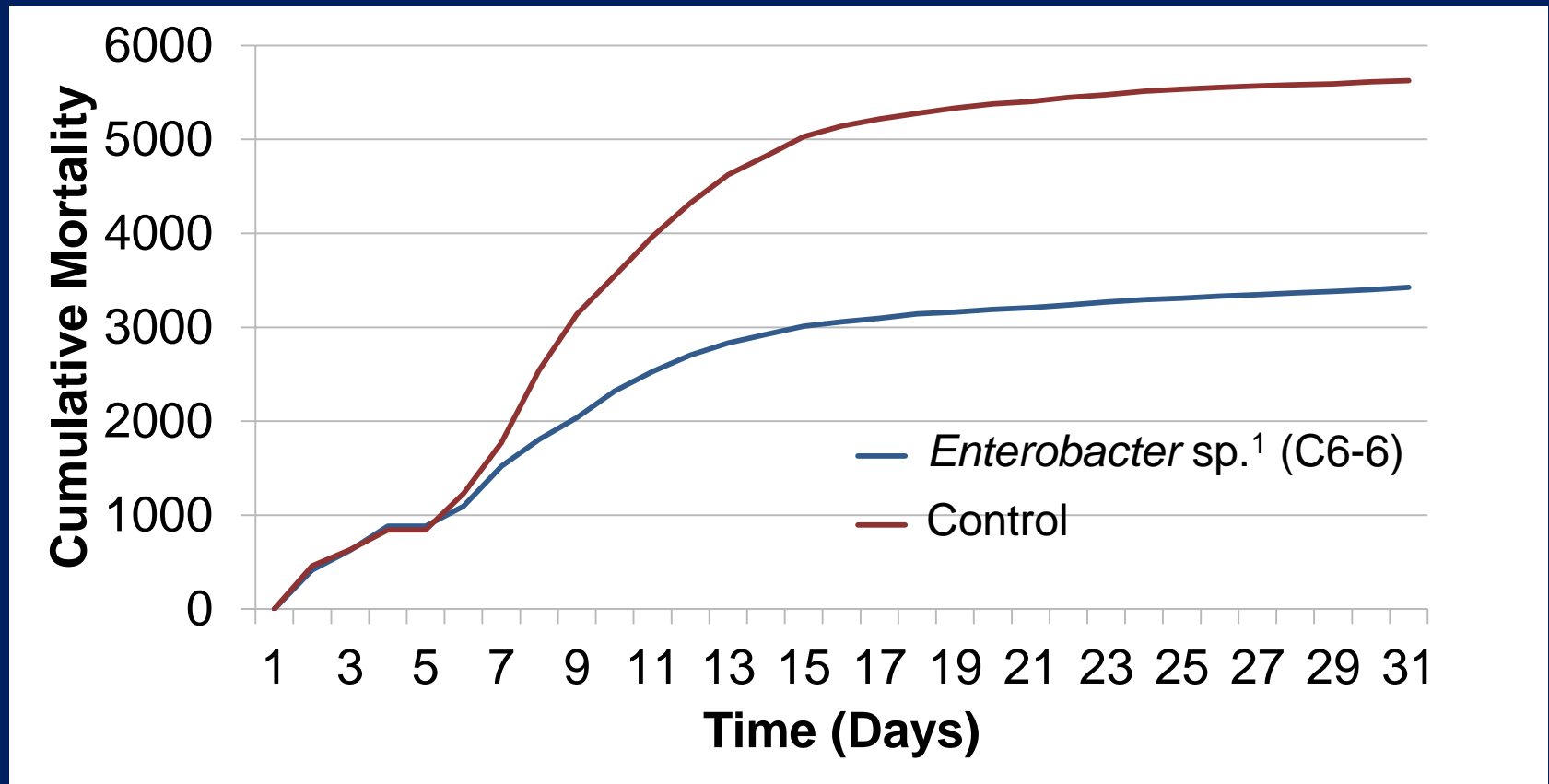
- *Enterobacter amnigenus* is an ubiquitous bacteria commonly found in the soil and water
- *Enterobacter* sp. PIC15 originally isolated from yellow catfish (*Pelteobagrus fulvidraco*)



Field Trials

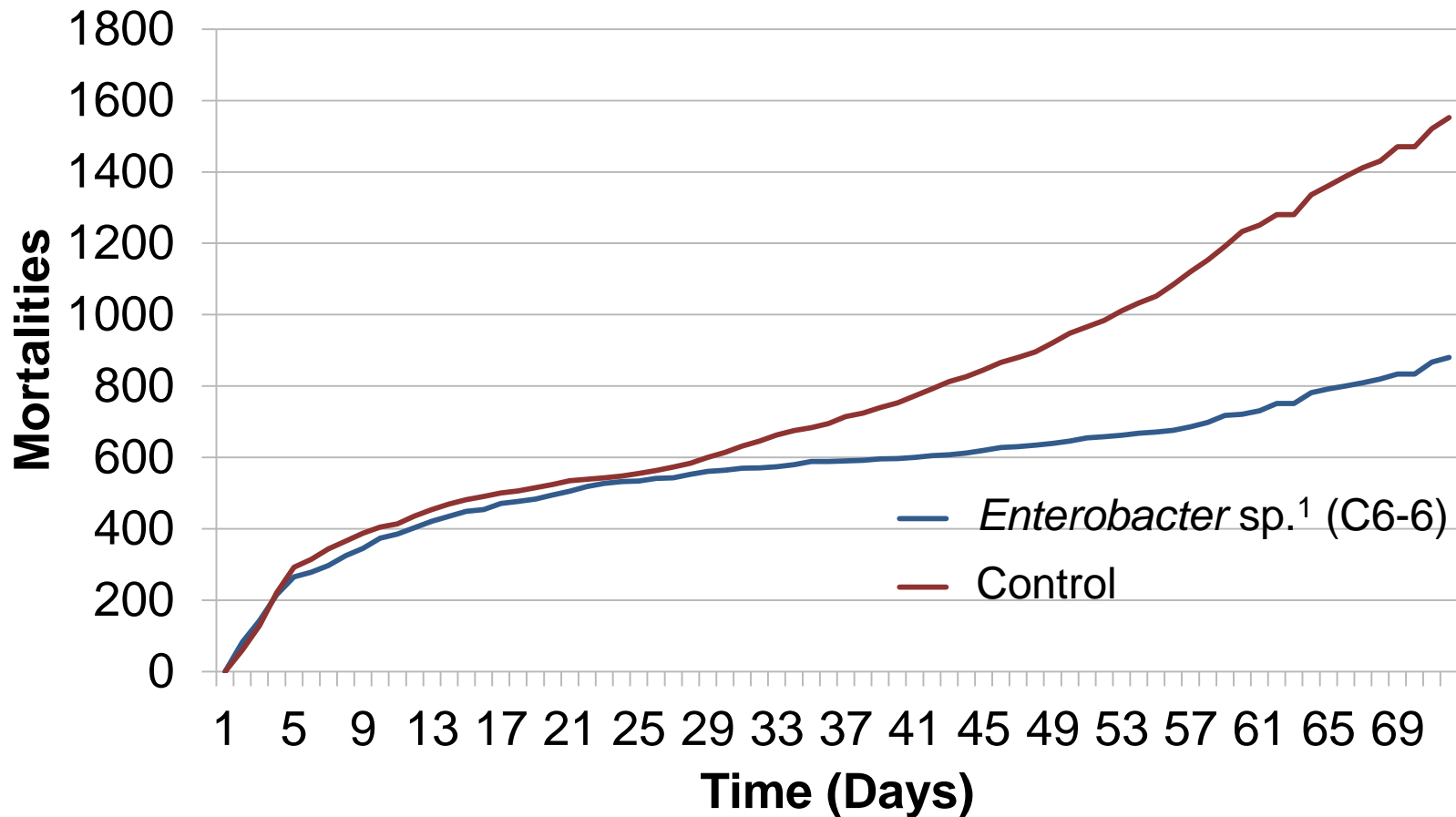
- *Enterobacter* sp.¹ (C6-6) provided to Utah Division of Wildlife Resources for field evaluations
 - Used at facilities experiencing *F. psychrophilum* outbreaks
 - Evaluated *Enterobacter* sp.¹ (C6-6) through several field trials

Field Trial 1



- ~ 38% decrease in total mortality from control group
- *F. psychrophilum* confirmed in both groups of fish

Field Trial 2



- ~ 43% decrease in total mortality from control group
- *F. psychrophilum* confirmed in both groups of fish

Discussion

- *Enterobacter* sp.¹ (C6-6) effective on a larger scale
 - Offers a relatively simple means of improving fish health (early life stages)



Summary (*Enterobacter* sp.)

- Two separate *Enterobacter* species acting similarly
 - Colonize the GI tract
 - Inhibit *F. psychrophilum* growth *in vitro*
 - Significantly reduce mortality due to *F. psychrophilum*
 - Non-pathogenic to rainbow trout and common in environment
- Other observations
 - Short doubling time ~ 30 min
 - Wide range of growth temperatures, 4 – 40 °C
 - Viability under storage conditions, -20, 4, 15 °C

Conclusions

Both *Enterobacter* species appear to represent a viable tool that can improve survival of fish infected with *F. psychrophilum*

- High potential (and need) for commercialization and application of these probiotics

It is possible that a combined approach using Probiotics at first feeding followed by Vaccination when fish are immunocompetent may dramatically reduce CWD problems

Acknowledgments

Funding

USDA

WSU / UI Aquaculture Initiative

McNair Achievement Program and Graduate Assistantship

Collaborators

- Christine Swan, Wade Cavender, and Chris Wilson, Utah Division of Wildlife Resources
- Devendra Shah, Washington State University
- Timberly Maddox, Nicole Lindstrom, Kurt Eversman, Matt Albert

Lab Support

- Mark Polinski
- Amy Long
- Tyson Fehringer