

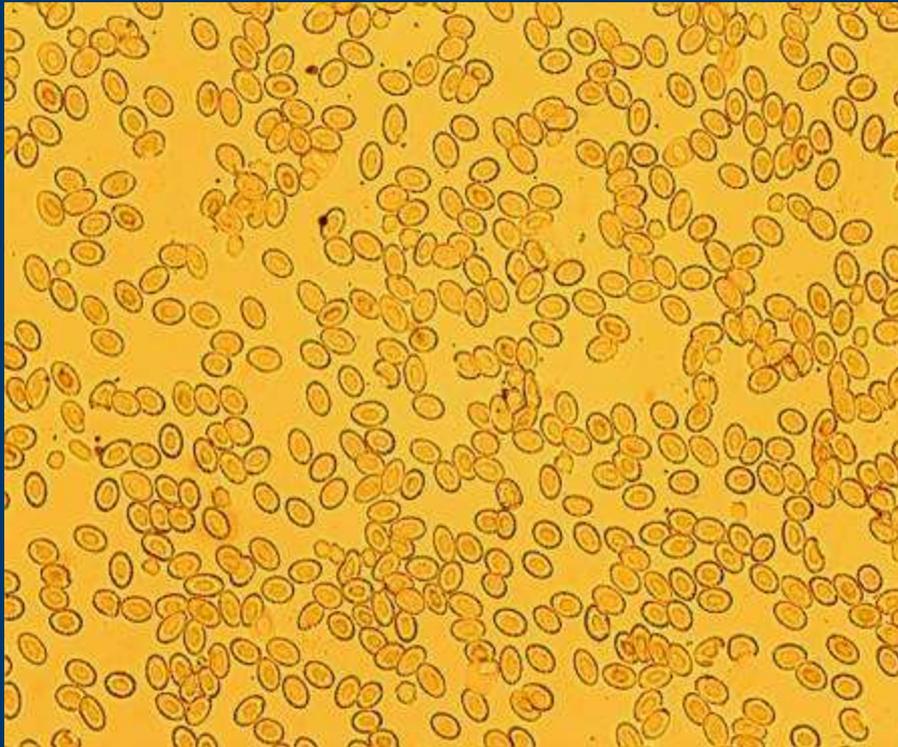


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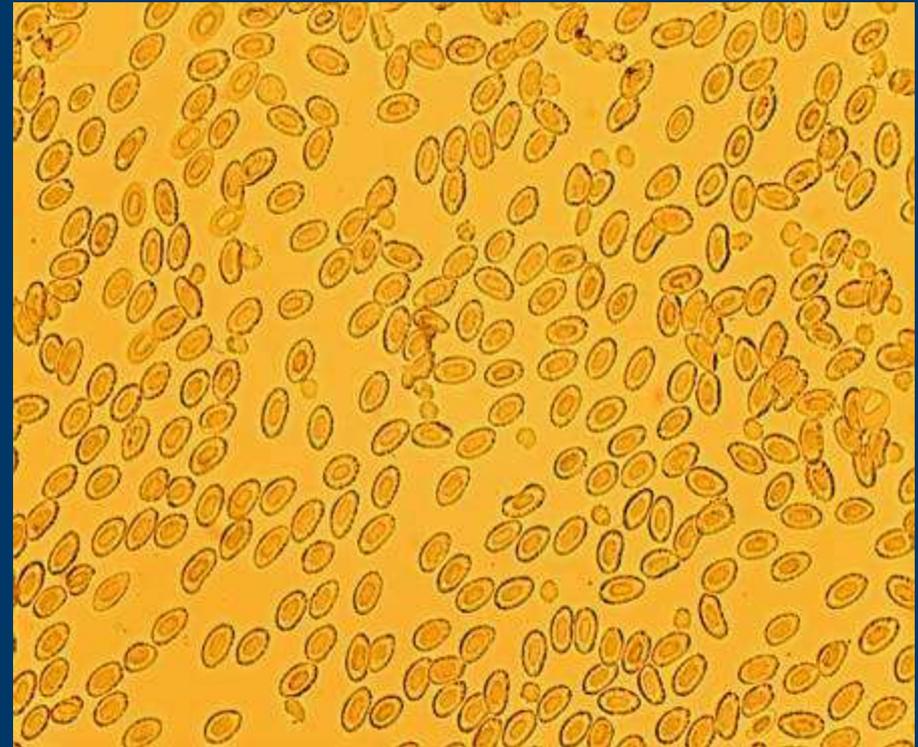
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**[gofishbc.com](http://gofishbc.com)**

# *A new method to confirm triploidy rates in salmonids*



Diploid



Triploid



# Outline

- Current FFSSBC production and sampling
- Difference of Imaging software (ImageJ) and Flow Cytometry
- Example of imaging software sample analysis



# *FFSBC Production and Sampling*

- Currently release 3 million triploid fish throughout the province of British Columbia
- Combination of hydrostatic pressure and heat shock since 1995
- Sampling to confirm triploid success at 0.05% of total release size



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# *Imaging Software*

## Pro's

- Basic Lab equipment
- No fish transfer
- Timeline extended
- Cost less per sample

## Con's

- New unknown process
- Sample Quality



# *Imaging Software*

- Blood smear
- Picture 200x magnification
- Upload pictures into software and run
- Output into datasheet



# *Flow Cytometry*

## Pro's

- Proven method
- Familiar with process

## Con's

- Time sensitive
- Shipping fish
- More expensive
- Chemical disposal
- Mutagen / Carcinogen

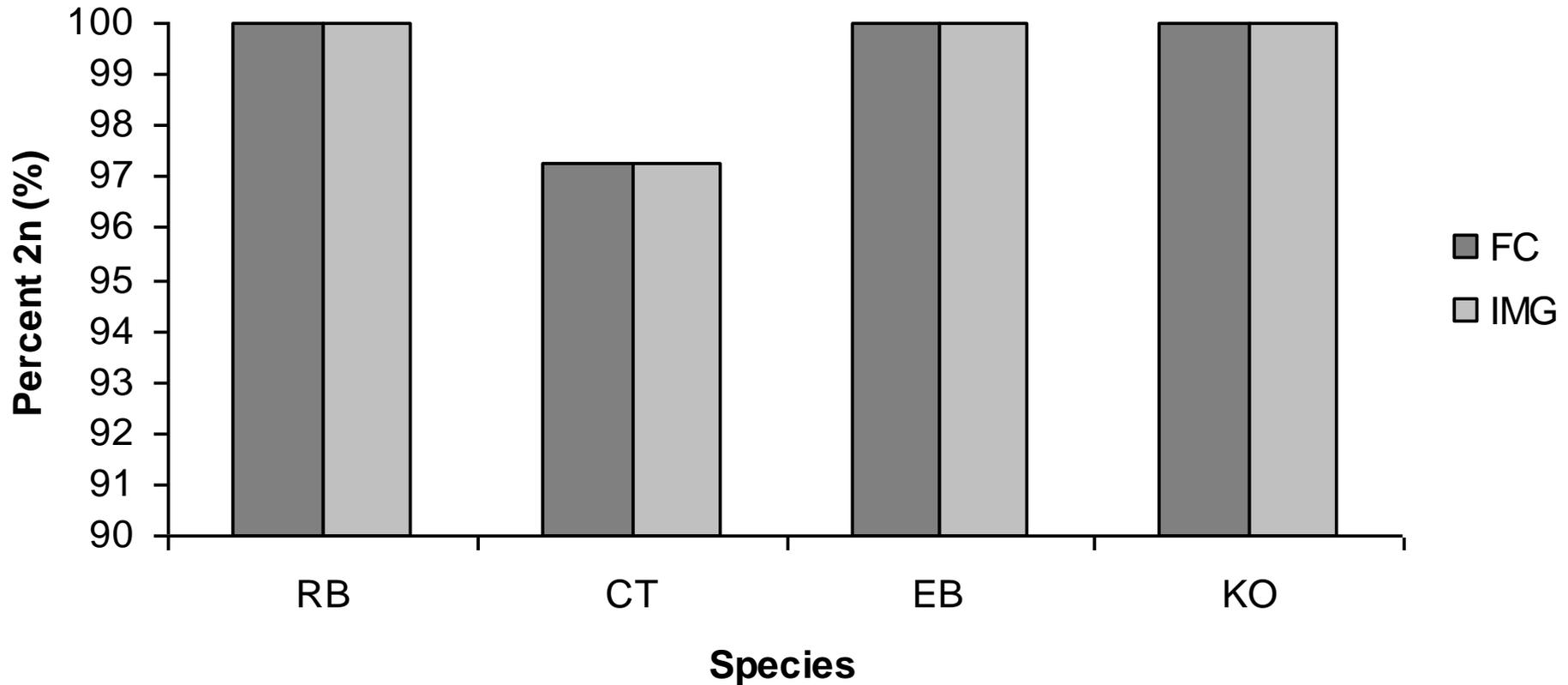


# *Flow Cytometry*

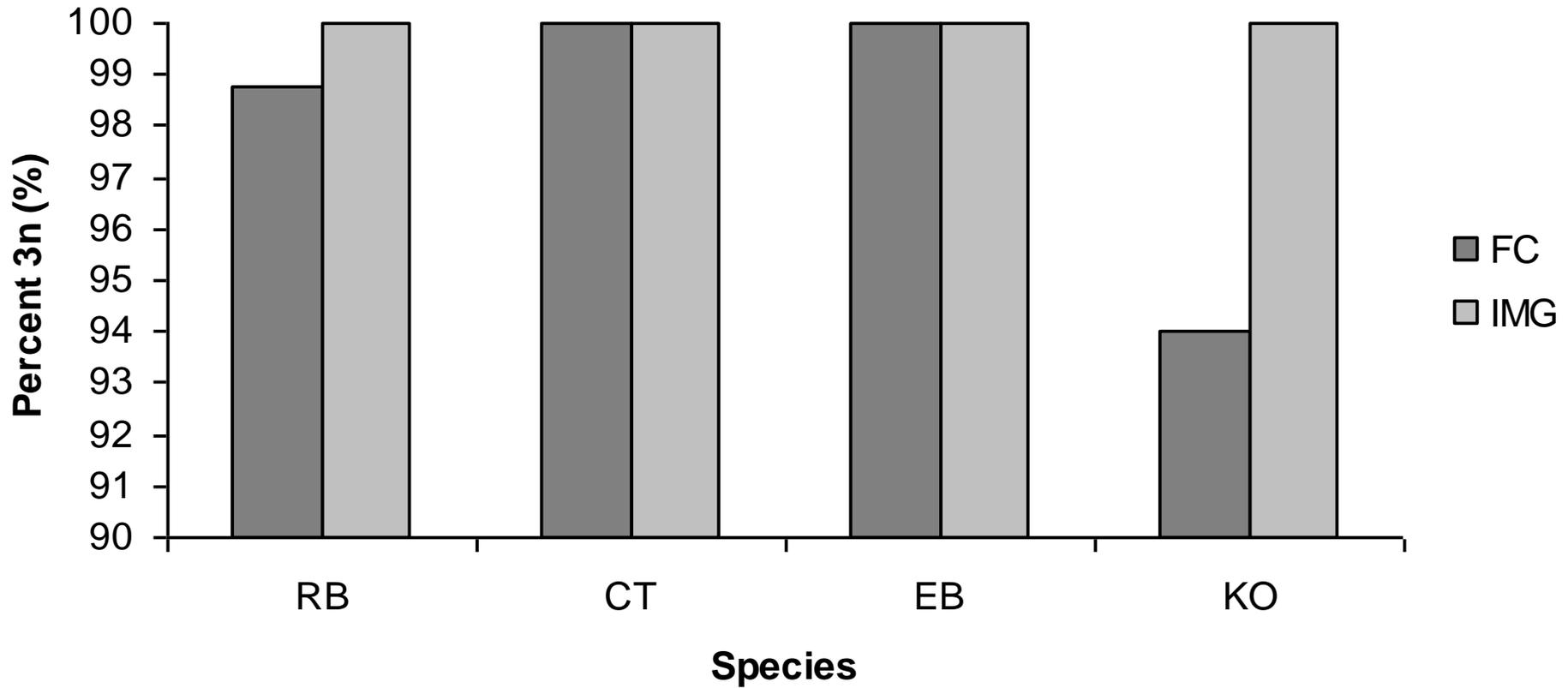
- Hold fish onsite, lethal sample
- Apply Nuclear Stain, Preserve
- Send to contractor
- Chemical disposal



# Comparison of Methods

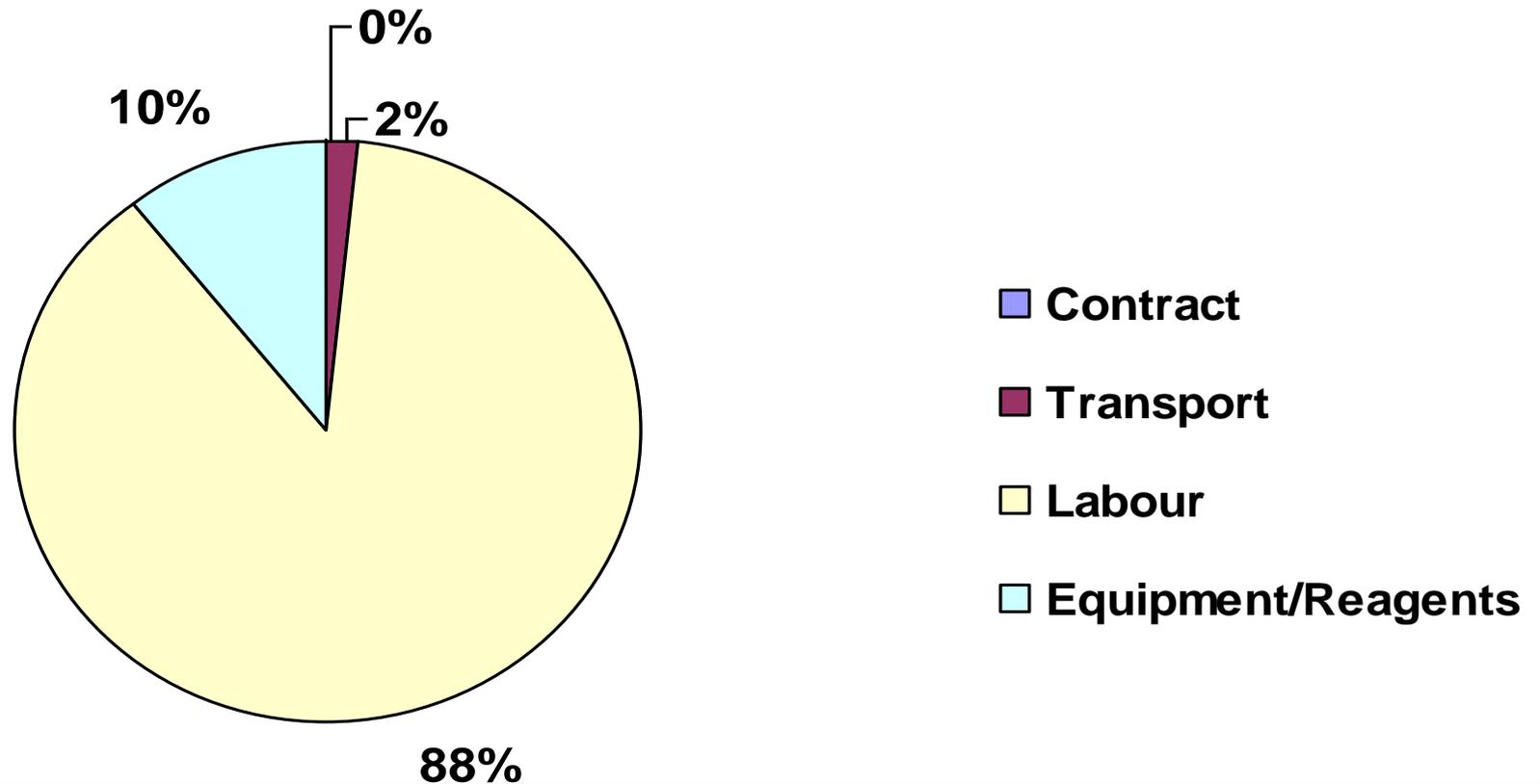


# Comparison of Methods

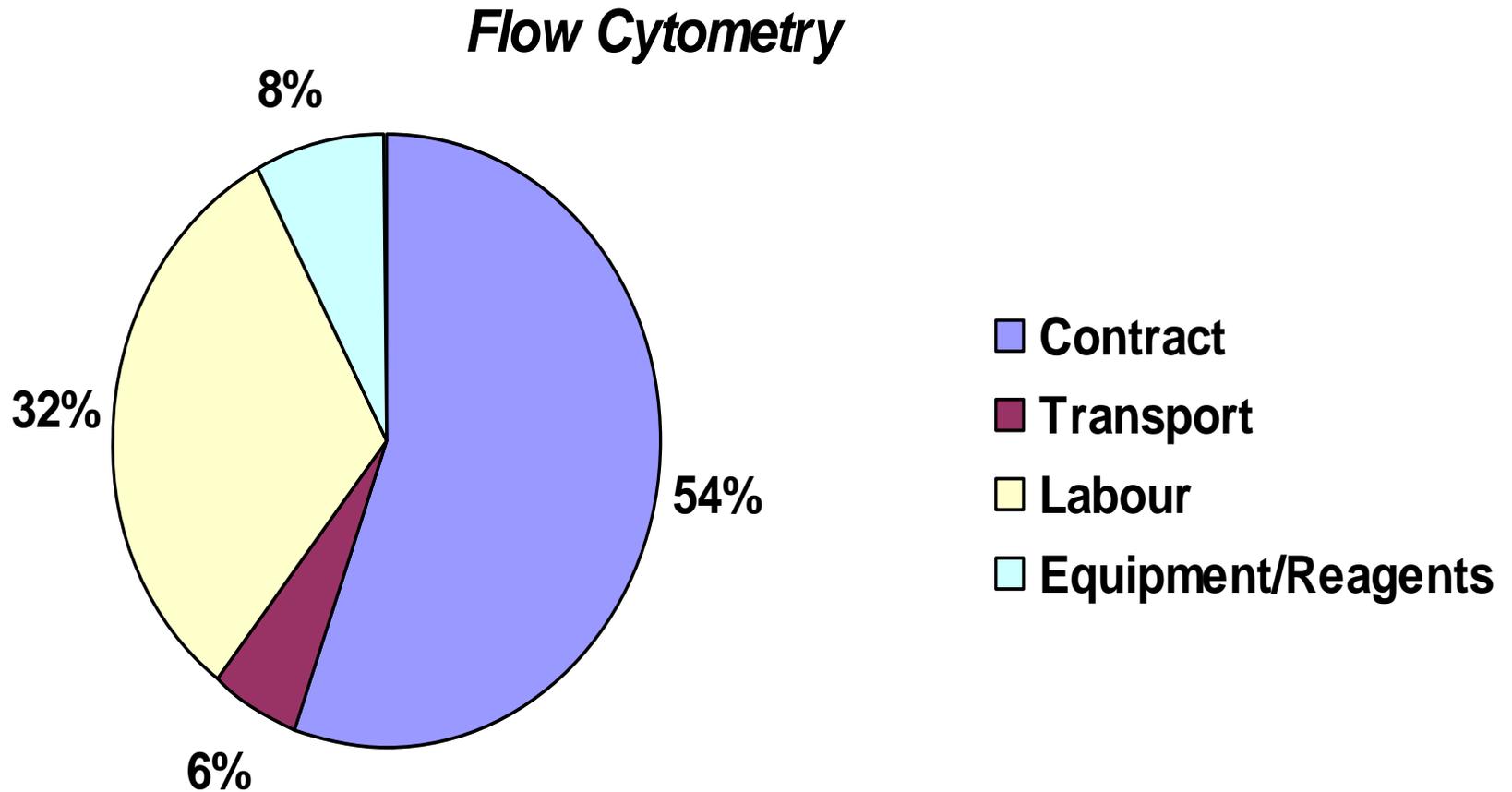


# Comparison of Methods

## Imaging



# Comparison of Methods



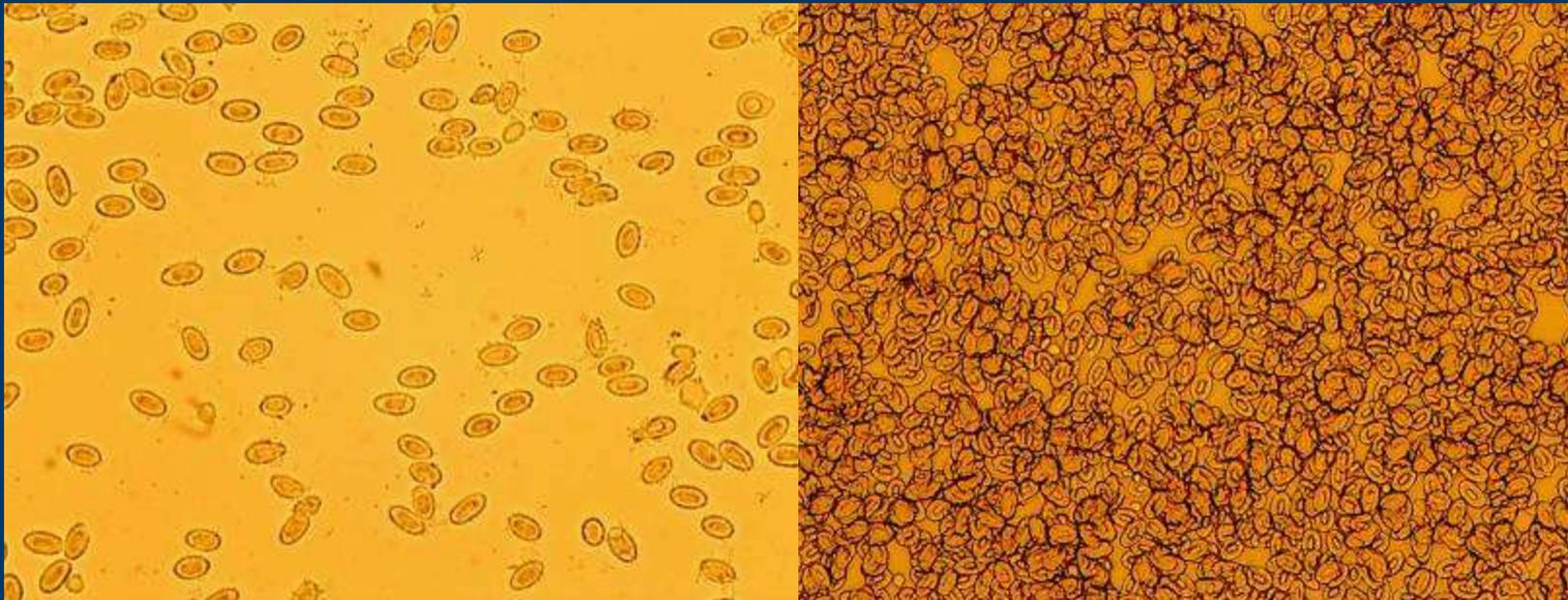
# Costing

- Based on 1900 samples
- Imaging Cost \$3.29 / sample
- Flow Cytometry \$6.40 / sample



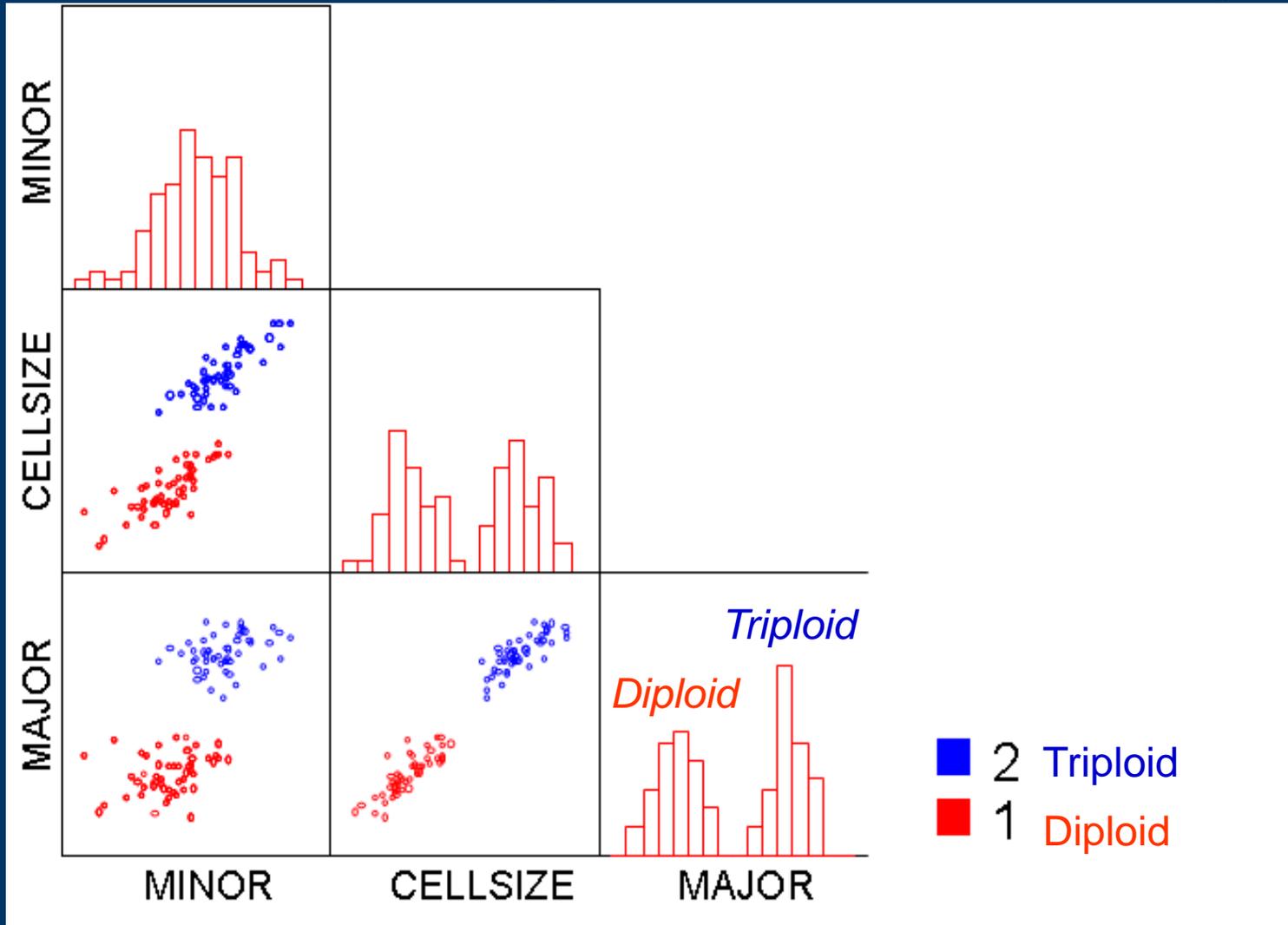
# *Imaging Software*

- Parameters
- Manipulation, measurement
- Organizing output, testing

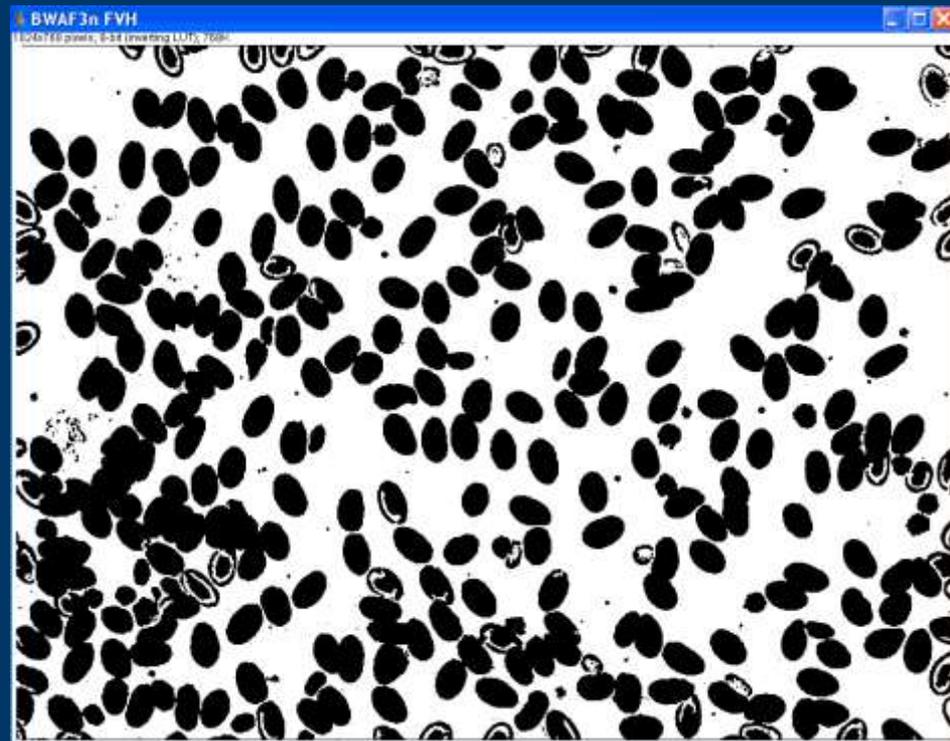
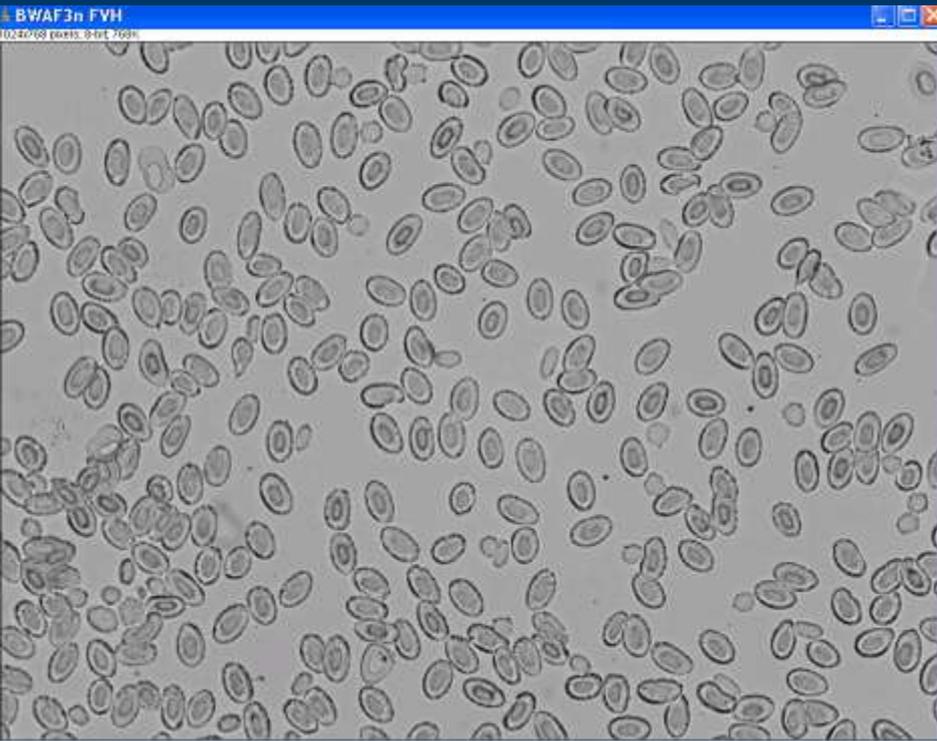


# Parameters

- Cell area, major/minor axis, circularity

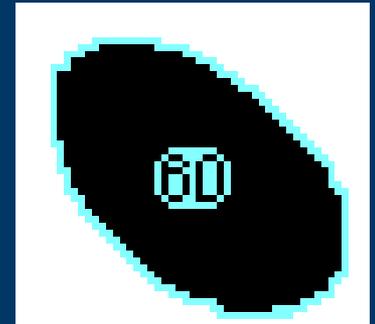
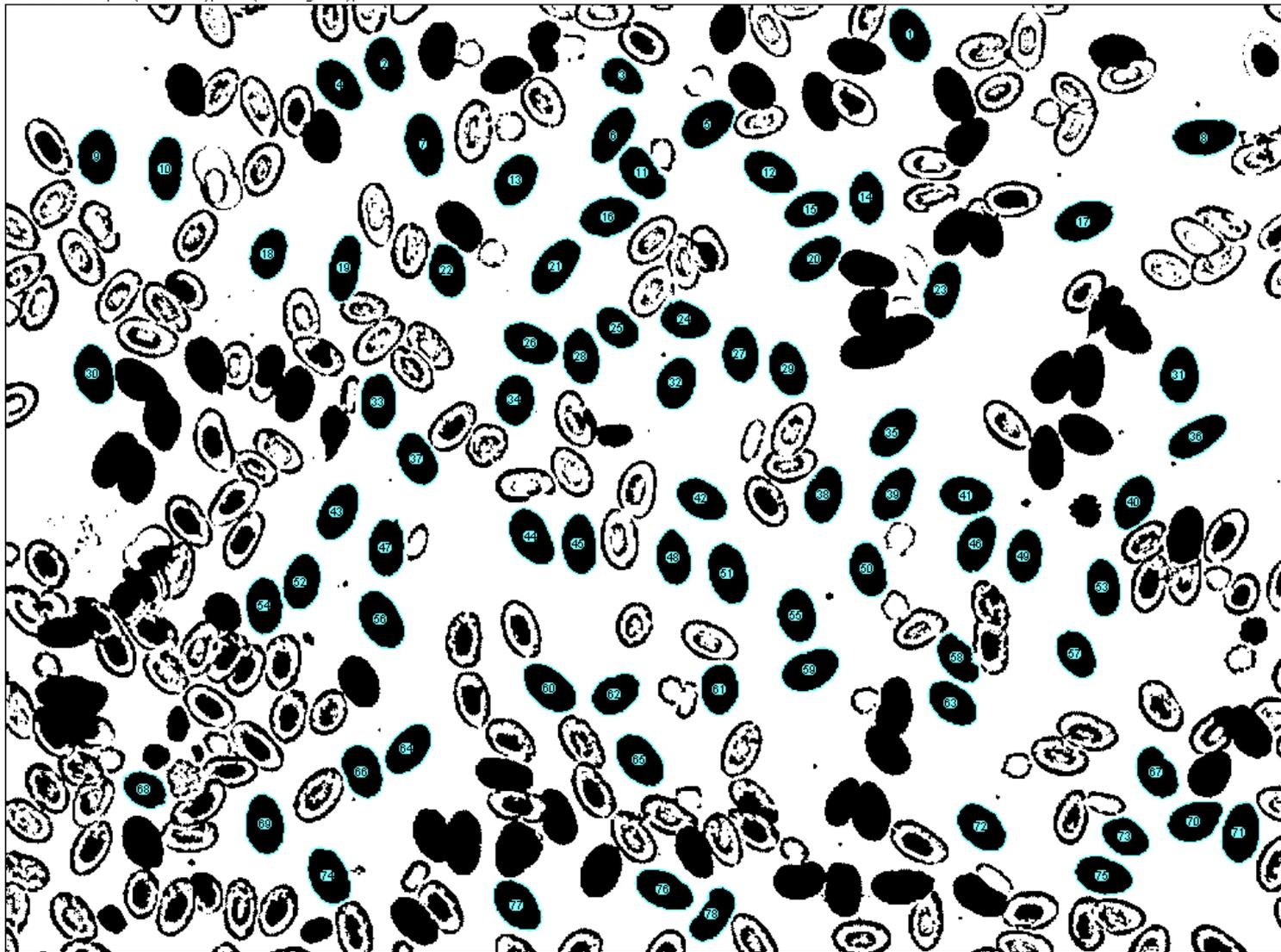


# *ImageJ Manipulation*



# ImageJ Manipulation

419.67x314.75 µm (1024x768); 8-bit (Inverting LUT); 768K



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# ImageJ Manipulation

- Output into spreadsheet
- Sort by major axis identify outliers
- Check individual cell measurements

YEAR	CELLID	STRAIN\$	AREA	MAJOR	MINOR	CIRC	FISHNO	PLOIDY\$
2011	1	BW	187.282	19.064	12.508	0.846	1	3n
2011	2	BW	190.641	18.669	13.002	0.814	1	3n
2011	3	BW	191.313	19.418	12.545	0.82	1	3n
2011	4	BW	207.438	19.94	13.246	0.825	1	3n



# *Analyzing*

- Import data from both genotypes
- Sort by major axis identify outliers
- Categorizes the sample based on the major axis measurement, 2 cluster analysis



# Analysis

- 2 cluster analysis using the long axis

Fish #		1		2		3	
		Freq	Percent	Freq	Percent	Freq	Percent
Cluster	1	0	0.00%	0	0.00%	0	0.00%
	2	56	100.00%	22	100.00%	83	100.00%
	Combined	56	100.00%	22	100.00%	83	100.00%



# Flowchart

Blood sample

<20 measurements

>20 measurements

Nuclei Measurements



2n/3n

Software to classify

Error or 2n classification then use Nuclei measurement

2n/3n



# Questions



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